Deacetylation of Fungal Exopolysaccharide Mediates Adhesion and Biofilm Formation

Mark J. Lee, Alexander M. Geller, Natalie C. Bamford, Hong Liu, Fabrice N. Gravelat, Brendan D. Snarr, François Le Mauff, José Chabot, Benjamin Ralph, Hanna Ostapska, Mélanie Lehoux, Robert P. Cerone, Stephanie D. Baptista, Evgeny Vinogradov, Jason E. Stajich, Scott G. Filler, P. Lynne Howell, Donald C. Sheppard

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ABSTRACT The mold Aspergillus fumigatus causes invasive infection in immunocompromised patients. Recently, galactosaminogalactan (GAG), an exopolysaccharide composed of galactose and N-acetylgalactosamine (GalNAc), was identified as a virulence factor required for biofilm formation. The molecular mechanisms underlying GAG biosynthesis and GAG-mediated biofilm formation were unknown. We identified a cluster of five coregulated genes that were dysregulated in GAG-deficient mutants and whose gene products share functional similarity with proteins that mediate the synthesis of the bacterial biofilm exopolysaccharide poly-(1-6)-N-acetyl-D-glucosamine (PNAG). Bioinformatic analyses suggested that the GAG cluster gene agd3 encodes a protein containing a deacetylase domain. Because deacetylation of N-acetylgalactosamine residues is critical for the function of PNAG, we investigated the role of deacetylation in fungal biofilm formation. Agd3 was found to mediate deacetylation of GalNAc residues within GAG and render the polysaccharide polycationic. As with PNAG, deacetylation is required for the adherence of GAG to hyphae and for biofilm formation. Growth of the Δagd3 mutant in the presence of culture supernatants of the GAG-deficient Δagd3 mutant rescued the biofilm defect of the Δagd3 mutant and restored the adhesive properties of GAG, suggesting that deacetylation is an extracellular process. The GAG biosynthetic gene cluster is present in the genomes of members of the Pezizomycotina subphylum of the Ascomycota including a number of plant-pathogenic fungi and a single basidiomycete species, Trichosporon asahii, likely a result of recent horizontal gene transfer. The current study demonstrates that the production of cationic, deacetylated exopolysaccharides is a strategy used by both fungi and bacteria for biofilm formation.

IMPORTANCE This study sheds light on the biosynthetic pathways governing the synthesis of galactosaminogalactan (GAG), which plays a key role in A. fumigatus virulence and biofilm formation. We find that bacteria and fungi use similar strategies to synthesize adhesive biofilm exopolysaccharides. The presence of orthologs of the GAG biosynthetic gene clusters in multiple fungi suggests that this exopolysaccharide may also be important in the virulence of other fungal pathogens. Further, these studies establish a molecular mechanism of adhesion in which GAG interacts via charge-charge interactions to bind to both fungal hyphae and other substrates. Finally, the importance of deacetylation in the synthesis of functional GAG and the extracellular localization of this process suggest that inhibition of deacetylation may be an attractive target for the development of novel antifungal therapies.

Aspergillus fumigatus is an opportunistic mold that causes invasive infections in immunosuppressed patients. Despite antifungal treatment with the currently available antifungal agents, the mortality of invasive aspergillosis (IA) remains between 50 and 95% (1), highlighting the urgent need for new, effective therapeutic agents for this disease. Identifying and targeting virulence factors unique to this fungus is one approach to the development of novel treatments for invasive aspergillosis.

Galactosaminogalactan (GAG) is an exopolysaccharide recently identified in A. fumigatus that plays a number of roles in the pathogenesis of IA (2, 3). GAG is a linear heteropolymer composed of α-1,4-linked galactose and N-acetylgalactosamine (GalNAc) that is bound to the outer cell wall and found within the extracellular matrix of biofilms of Aspergillus species (4, 5). Cell-wall-associated GAG mediates a number of virulence-associated traits, including adherence to host cells and other substrates, bio-

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Address correspondence to Donald C. Sheppard, don.sheppard@mcgill.ca.

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film formation, masking of β-1,3-glucans from immune recognition, and resistance to neutrophil extracellular traps (2, 6). However, the biosynthetic pathways underlying GAG synthesis and the molecular mechanisms by which GAG mediates these virulence traits remain poorly understood.

A number of pathogenic bacteria also produce exopolysaccharide composed of N-acetylhexosamines, most commonly β-1,6-linked N-acetylglucosamine (GlcNAc). This exopolysaccharide, known as polysaccharide intercellular adhesion (PIA) in Staphylococcus aureus (7) or poly-(β1-6)-N-acetyl-d-glucosamine (PNAG) in Escherichia coli (8), mediates many of the same virulence properties as GAG does, including adherence of the organism to both biotic and abiotic surfaces, resistance to neutrophil killing, and masking of pathogen-associated molecular patterns (PAMPs) (8–11). The mechanisms governing the syntheses of these bacterial exopolysaccharides have been well characterized and are mediated by the gene products of the ica or pga operons, respectively (Fig. 1A and B and Fig. 2). Briefly, PIA/PNAG is synthesized and transported.

FIG 1 Bacterial polysaccharide biosynthetic operons and the putative GAG biosynthetic gene cluster. (A to C) Schematic of the ica operon responsible for the synthesis of polysaccharide intercellular adhesion (PIA) (A), pga operon responsible for the synthesis of PNAG [poly-(β1-6)-N-acetyl-d-glucosamine] (B), and the putative GAG gene cluster (C). (D) Heatmap showing differential gene expression of the A. fumigatus MedA and StuA regulatory mutants compared to wild-type A. fumigatus, highlighting the coregulation of the genes in the GAG biosynthesis gene cluster. Fold induction is shown in red (upregulation), green (downregulation), black (no change), and gray (missing data point). Locus ID, locus identification.
across the membrane and extruded extracellularly by the action of a glycosyltransferase complex composed of either IcaA/D or PgaC/D, respectively. The nascent polysaccharide undergoes partial deacetylation of GlcNAc residues by IcaB/PgaB, resulting in interspersed glucosamine (GlcN) residues (11–13). In Gram-negative bacteria, the partially deacetylated polymer is then transported across the outer membrane by PgaA (14). Under acidic conditions, these GlcN residues are protonated, conferring a positive charge on the mature polysaccharide, and thereby mediating adherence to negatively charged surfaces including host cells and the organism itself, as well as enhancing resistance to cationic molecules such as aminoglycoside antibiotics and antimicrobial peptides (15). Loss of deacetylase activity results in the formation of a fully acetylated polysaccharide that cannot adhere to the organism or other substrates, and which is shed into the culture supernatant, rendering it nonfunctional (11, 16).

Although the type of hexosamine and linkages differ between PIA/PNAG and GAG, we hypothesized that the biosynthetic pathways for these glycans are similar and that partial deacetylation of GAG is also required for its function. Comparative transcriptome studies identified a cluster of genes in *A. fumigatus* that is similar in composition to the bacterial exopolysaccharide operons. This cluster encompasses the *uge3* gene, encoding a glucose 4-epimerase that is required for GAG biosynthesis (2, 17). Molecular and biochemical studies revealed that, as with bacterial exopolysaccharide, GAG is partially deacetylated. This deacetylation is mediated by Agd3, which is encoded by another gene within the GAG biosynthetic gene cluster. Extracellular Agd3 activity was required for binding of GAG to the hyphal cell wall, biofilm formation, masking of β-1,3-glucan, and full virulence. Bioinformatic analysis of fungal genomes revealed the presence of the GAG biosynthetic gene cluster within the genome of a number of other filamentous ascomycetes. The complete cluster and evidence for GAG production was also found within the basidiomycete *Trichosporon asahii*, likely as a consequence of a recent horizontal gene transfer event.

**RESULTS**

Identification of a gene cluster encoding putative GAG biosynthetic proteins. Previously, comparative transcriptomic analysis of two transcription factor mutants, ΔstuA and ΔmedA mutants, identified *uge3*, a gene encoding a bifunctional epimerase, as required for GAG synthesis (2, 17). A more detailed analysis of this data set revealed that the *uge3* gene is flanked by four other coregulated genes (Fig. 1C and D). These genes include genes predicted to encode a glycosyltransferase (gtb3), polysaccharide deacetylase (agd3), polysaccharide hydrolase (ega3), and spherulin-4-like protein (sp3h). The predicted functions of these proteins suggest a model for exopolysaccharide synthesis analogous to the bacterial ICA/PGA systems (Fig. 2A). In this model, UDP-galactose and UDP-GalNAc are synthesized by the activity of the epimerase Uge3 and then polymerized and extruded from the cell by the putative transmembrane glycosyltransferase Gtb3. As with bacterial exopolysaccharide systems, the function of the putative glycosyl hydrolase within the fungal system is unclear, but a recent study from our group suggests that it may be required for polymer modification and remodeling (18). Importantly, the final step in this putative pathway is predicted to be the partial deacetylation of N-acetylgalactosamine residues within the nascent polymer by the deacetylase Agd3 to generate a polycationic glycan that adheres to the hyphal and other surfaces. To begin to validate this model, we therefore sought to determine the role of Agd3 in GAG deacetylation and function.

**Agd3 is predicted to contain a polysaccharide deacetylase domain.** Bioinformatic analysis of Agd3 suggests that it is a multidomain extracellular protein including a predicted signal peptide with a cleavage site between Cys18 and Thr19 (19, 20), suggesting that Agd3 is secreted as an extracellular protein (Fig. 2B). Structure prediction server Phyre2 analysis of Agd3 predicted several distinct domains and extended regions of low complexity (21). The N-terminal region of the protein from residues 38 to 129 contains a serine-rich region that is predicted to be structurally disordered. When Agd3 lacking the signal peptide was analyzed using the structure prediction server Phyre2, the top hits (>95% confidence) for residues 509 to 726 were deacetylase domains of members of carbohydrate esterase family 4 (CE 4) (3A). The CE 4 family contains a large number of metal-dependent polysaccharide deacetylases, which have been shown to remove N- or O-linked acetate groups from chitin, peptidoglycan, acetylxylan, and poly-β-1,6-N-acetylglucosamine (12, 22–24). CE 4 domains adopt a (β/α) fold with five canonical amino acid motifs that are important for active site formation and activity (12, 23). The structural prediction for Agd3 contains only a partial CE 4 fold starting at the second active site motif, which usually occurs at the end of the third β-strand of the barrel (Fig. 2B).

Using the whole protein, Phyre2 was unable to predict any function-associated motifs for residues 140 to 509 (25). However, when submitted to Phyre2 in isolation, residues 147 to 234 were predicted (86.2% confidence) to be similar to the flavodoxin-like fold present in *Thermotoga maritima* class I enzyme amido-transferase (GATase) (PDB code 1O1Y). Class I GATases are defined by a conserved Cys-His-Glu catalytic triad that is important for the reductase activity (25). These residues could not be identified in the Agd3 model, which contained only 88 of the 140 to 180 residues typically found in this fold. This domain is followed by a region of low structural complexity, as residues 243 to 300 were predicted to be largely disordered by both Phyre2 and GlobPlot. When residues 200 to 500 were submitted, a partial (β/α) barrel was identified between residues 392 and 468 with 75.3% confidence. This region may represent the first β/α repeats of the CE 4 domain.

The metal coordinating triad (Asp-His-His) is conserved in CE 4 enzymes and is localized to two of the five canonical linear motifs (MTs). The MT1, DD and DXD motifs contain the coordinating aspartic acid, as well as the catalytic base (Fig. 2C). In Agd3, MT1 is most likely D377 and D378, which are highly conserved in Agd3 orthologs. These residues are located outside the predicted CE 4 domain and slightly upstream of the partial (β/α) barrel motif. MT2 contains the metal coordinating histidines. In Agd3, H510 and H514, which are highly conserved and align with active site residues of known polysaccharide deacetylases using ClustalW, probably constitute MT2 (2C). The linear distance between MT1 and MT2 is larger than that found in most CE 4 family members and is predicted to be a region of low structural complexity. Similar insertions between motifs are present in *Helicobacter pylori* PgdA and *Staphylococcus epidermidis* IcaB, where they are important for homomultimerization and membrane association, respectively (26, 27). Circular permutations of the CE 4 motifs have also been reported in multiple members (23, 26). Based on these findings, Agd3 has a unique CE 4 domain compo-
sition that may have prevented accurate modeling of the entire domain by Phyre2, making the exact domain boundaries unclear. Collectively, these in silico analyses suggest that Agd3 contains functional domains similar to known bacterial polysaccharide deacetylases, and therefore Agd3 may mediate the deacetylation of GAG.

Deletion of agd3 blocks deacetylation of GAG. To determine whether GAG is partially deacetylated, the presence of primary amines in purified GAG was quantified by a chemical approach using trinitrobenzene sulfonate (TNBS) (28). This analysis identified that the presence of primary amines consistent with the presence of non-acetylated galactosamine (GalN) in GAG purified from wild-type *A. fumigatus* (Fig. 3A). The presence of GalN residues within purified GAG was also confirmed by nuclear magnetic resonance (NMR) analysis (Fig. 3B). Taken together, these data suggest that GAG undergoes partial deacetylation.

To determine whether Agd3 activity is required for GAG deacetylation, a Δagd3 deletion mutant strain was constructed. Deletion of agd3 resulted in the production of GAG that was devoid of primary amines and lacked GalN as detected by NMR analysis (Fig. 3A and B). Deletion of agd3 did not significantly affect the production of total secreted GAG (Fig. 3C) or affect the expression of other genes in the GAG biosynthetic gene cluster under all growth conditions tested (Fig. 3D). As previously reported for the deletion of uge3, loss of Agd3 did not affect susceptibility to antifungal agents or cell wall stressors, including caspofungin and nikkomycin (see Fig. S1 in the supplemental material). Complementation of the Δagd3 mutant with a wild-type allele of
agd3 restored deacetylation to wild-type levels. These data suggest that Agd3 is required for the deacetylation of GAG.

**Deletion of agd3 is associated with the loss of adherence and changes in the cell wall morphology.** To examine the role of deacetylation and significance of the resulting cationic charge on the functional properties of GAG, the Δagd3 mutant was compared with the wild-type parent and agd3 complemented strains for a variety of GAG-dependent phenotypes. Deletion of agd3 was associated with a loss of adherence to negatively charged tissue culture-treated surfaces (Fig. 4A). However, consistent with the hypothesis that GAG-mediated adherence is dependent on charge-charge interactions, the Δagd3 mutant retained the ability to form wild-type biofilms on positively charged poly-D-lysine-coated plates (Fig. 4A). Thus, deacetylation of GAG mediates adherence to negatively charged surfaces, but not to positively charged surfaces.

The loss of Agd3 resulted in the loss of detectable cell wall-associated GAG as measured by direct immunofluorescence with the GaINAc-specific soybean agglutinin (SBA) lectin (Fig. 4B). These findings were also confirmed by scanning electron microscopy, which demonstrated a loss of the GAG-dependent cell wall decorations (Fig. 4C). Consistent with this loss of cell wall-associated GAG, hyphae of the Δagd3 mutant displayed increased exposure of β-1,3-glucan as detected by enhanced binding of re-
combinant Fc-dectin-1 by fluorescence microscopy (Fig. 4D; see Fig. S2 in the supplemental material). The phenotype of the Δagd3 mutant observed in these assays was indistinguishable from the GAG-deficient Δuge3 mutant (2). Collectively, these data suggest that Agd3-mediated deacetylation of GAG is required for adherence of the polysaccharide to the fungal cell wall and mediates adherence of hyphae to positively charged surfaces.

De-N-acetylated GAG confers a positive charge on the hyphal surface. To confirm that the presence of deacetylated GAG alters the surface charge of A. fumigatus hyphae, the ability of hyphae to bind negatively charged Sephadex beads was quantified (29). When coincubated with anionic Sephadex beads, there was a greater binding of negatively charged beads by wild-type A. fumigatus or by the agd3 complemented strain than by the Δagd3 mutant (Fig. 5). Binding of anionic beads by hyphae of the Δagd3 mutant was similar to the binding of these beads by the completely GAG-deficient Δuge3 mutant. Thus, deacetylation is required to render GAG cationic and mediates adhesion to both the hyphal cell wall and other surfaces.

Deletion of agd3 attenuates virulence. To confirm that deacetylation of GAG plays a role in pathogenesis, the virulence of the Δagd3 mutant was compared to the wild-type Af293 strain and the agd3 complemented strain in a leukopenic murine model of invasive aspergillosis (Fig. 6A) (2, 30). Consistent with the in vitro findings of impaired GAG function, the Δagd3 mutant was hypovirulent compared to wild-type A. fumigatus or the agd3 complemented strain. Histopathologic examination of the lungs of mice infected with the Δagd3 mutant revealed no difference in the appearance of fungal lesions in mice infected with the wild-type or Δagd3 mutant strains (Fig. 6B). However, determination of fungal burden by quantitative PCR (Fig. 6C) or pulmonary galactomannan content (Fig. 6D) revealed a lower fungal burden in the lungs of mice infected with the Δagd3 mutant than in mice infected with the wild-type parent strain, as was previously reported with the Δuge3 mutant strain. Consistent with the lower fungal burden observed in mice infected with the Δagd3 mutant, lower pulmonary injury was observed in these animals as measured by lactose dehydrogenase activity in bronchoalveolar lavage fluid (Fig. 6E). Thus, deacetylation of GAG is required for full virulence of A. fumigatus.
Mann-Whitney test as indicated by the § symbol. 

strain and those infected with either the fungal burden or lung injury in mice infected with wild-type Af293
ans plus interquartile ranged (error bars). There was a significant difference in
with the indicated strains. There were 8 mice in each group. Values are medi-
dehydrogenase activity in the bronchoalveolar lavage fluid of mice infected
There were 8 mice in each group. (E) Pulmonary injury as measured by lactose
strains, as measured by determination of pulmonary galactomannan content. 

FIG 6 Agd3 is required for full virulence in a mouse model of invasive

A

B

C

D

E

FIG 7 Culture filtrates from the Δuge3 mutant complement the defects in
adherence and cell wall morphology of Δagd3 mutant. (A) Biofilm formation
by the indicated strains grown alone or in coculture. After the biofilm was
washed, adherent biofilm was visualized by crystal violet staining. (B) Biofilm
formation by the indicated strains grown in the presence of culture filtrates
(C/F) from the Δuge3 or Δagd3 mutant. After the biofilm was washed, adher-
ent biofilm was visualized by crystal violet staining. (C) Scanning electron
microscopy visualization of cell wall morphology of the indicated strains
grown in the presence of culture filtrates from the Δuge3 mutant. The white
arrows indicate cell wall decorations associated with cell wall-bound GAG.

Restoration of biofilm formation by coculture of biofilm-deficient strains. Since the Δagd3 mutant produces fully acety-
lated GAG but not Agd3, and the Δuge3 mutant produces Agd3 but not GAG, we hypothesized that these strains could complement one another to produce deacetylated GAG and form biofilms. As predicted, coculture of the Δuge3 and Δagd3 mutants resulted in the formation of biofilms that were indistinguishable from those produced by wild-type A. fumigatus (Fig. 7A). Further, growth of the Δagd3 mutant in the presence of culture filtrates from the Δuge3 mutant resulted in the formation of adherent biofilms (Fig. 7B) and restored the presence of GAG-associated cell wall decorations as detected by scanning electron microscopy (Fig. 7C). Growth of the Δagd3 or Δuge3 mutant in the presence of their own respective culture filtrates had no effects on adherence (Fig. 7B) or hyphal morphology (Fig. 7C).

To extend these findings, we also examined the ability of cell-free culture filtrates from both Δagd3 and Δuge3 mutants to generate deacetylated GAG in vitro in the absence of A. fumigatus hyphae. Culture supernatants of each mutant or a 1:1 mixture of both culture supernatants were incubated in anionic polystyrene enzyme immunoassay (EIA) plates to capture adherent, deacetylated GAG. Adherent, deacetylated GAG was then quantified using an anti-GAG antibody (3). A 1:1 mixture of culture supernatants of the Δagd3 and Δuge3 mutants resulted in detection of adherent GAG at a level similar to that recovered from culture supernatants of wild-type A. fumigatus (see Fig. S3 in the supplemental material), while no adherent GAG was detected in culture filtrates of the Δagd3 mutant or Δuge3 mutant alone. Collectively, these results support the proposed model of GAG biosynthesis in which GAG undergoes deacetylation by extracellular Agd3.
Agd3 is localized on the hyphal surface. The results of our biochemical and complementation studies suggest that Agd3-mediated deacetylation is an extracellular process. Consistent with these findings, a recent study identified Agd3 within the secretome of *A. fumigatus* (31). To determine whether Agd3 is also present within the cell wall, a red fluorescent protein (RFP)-tagged *agd3* allele was expressed under the native *agd3* promoter in *A. fumigatus* Af293. In this strain, while low levels of red fluorescence in the cell wall were observed, the signal was obscured by significant red autofluorescence of hyphae. To enhance visualization of RFP-tagged Agd3 (Agd3-RFP), indirect immunofluorescence using a fluorescein isothiocyanate (FITC)-tagged anti-RFP antibody was performed. Using this technique, low levels of Agd3-RFP were observed on the surfaces of hyphae (Fig. 8). Western blot analysis of culture supernatants and fungal biomass confirmed that although Agd3-RFP was found both in the culture supernatant and the fungal biomass, the majority of Agd3-RFP was associated with hyphae (see Fig. S4 in the supplemental material). Interestingly, from the fungal biomass, four discrete bands were detected using anti-RFP antibodies, while in the secreted fraction, only a single band was observed. The same pattern of bands was detected in multiple independent experiments. Liquid chromatography-mass spectrometry (LC-MS) analysis of all five of these bands confirmed that these bands contained Agd3-RFP. Further, the relative abundance of individual peptides did not vary significantly between these bands, suggesting that the differences in band migration are more likely due to differences in glycosylation rather than proteolytic cleavage.

The GAG biosynthesis gene cluster is widespread in *Ascomycetes* but present only in *Trichosporon asahii* among *Basidiomycetes*. Using an *in silico* approach, the GAG gene cluster sequences from *A. fumigatus* were queried against more than 250 publically available fungal genomes at NCBI Genomes and FungiDB (32). Orthologous genes of the GAG gene cluster present at the same locus were found in 28 fungal species. In the majority of these species, synteny of the cluster genes was observed, suggesting a common origin for this cluster in these fungal species (see Fig. S5 in the supplemental material). All of the species containing the GAG gene cluster belong to the *Pezizomycotina* subphylum of the *Ascomycota* with the exception of one member of the phylum *Basidiomycota*, *Trichosporon asahii* (Fig. 9). More than half of the fungal species that possess the GAG gene cluster have been described as pathogenic species (see Table S1 in the supplemental material). Of these species, 13 are plant pathogens, while only 3 are known pathogens to insects and animals, suggesting a possible role for GAG in the pathogenesis of plant fungal infections.

To elucidate the evolutionary relationship among these gene cluster-containing species, and specifically of Agd3 orthologs, we constructed a gene tree by aligning the amino acid sequences of Agd3 orthologs from all 28 gene cluster-containing species (Fig. 9) and constructed a maximum likelihood tree rooted by the copies from the early branching ascomycete and nematode-trapping fungus *Arthrobotrys oligospora*. While the relationship among the orthologs of Agd3 largely mirrors the evolutionary relationship of the respective species as established by the fungal tree of life (33, 34), we found that the *T. asahii* Agd3 ortholog was nested in the tree within members of the class Sordariomycetes (Fig. 9), suggesting that a recent horizontal gene transfer event may have occurred. Consistent with this hypothesis, a predicted mutator-like element (MULE) domain-containing protein was found within the GAG gene cluster of *T. asahii* (see Fig. S5 in the supplemental material). The MULE domain-containing proteins are class II DNA transposases (35) and are found in fungal species (36) and are commonly found in eukaryotes (37). Analysis of the sequences flanking the cluster did not reveal an identifiable tandem inverted repeat at the borders of the GAG synthetic cluster in *T. asahii* or any other mobile element signatures. It therefore seems most likely that the transposable element is no longer active and that only the transposase domain remains, while other features of the mobile element have degraded.

To ascertain whether the GAG cluster was functional in *T. asahii*, mature hyphae of this organism were stained with fluorescein-tagged SBA lectin to detect the production of GAG. Lectin staining suggests that *T. asahii* secretes GAG into the extracellular matrix (Fig. 10). GAG produced by *T. asahii* was less uniformly bound to the cell wall of hyphae compared with *A. fumigatus* and seemed to form a matrix between hyphae.

**DISCUSSION**

In the present study, we report a cluster of coregulated genes on chromosome 3 of *A. fumigatus* with composition and functions similar to those of bacterial operons encoding proteins required for exopolysaccharide synthesis. Characterization of one of the enzymes in this cluster demonstrates that, as with bacterial exopolysaccharide, deacetylation of GAG is required for GAG to adhere both to substrates and to the organism itself, and for full virulence. Further, these studies suggest that GAG-mediated adherence is largely a consequence of charge-charge interactions between the polycationic polysaccharide and negatively charged surfaces.

The results of this study illustrate important similarities between the Agd3 deacetylase and other multidomain CE 4 family members from bacteria. In *S. epidermidis*, deacetylation of PIA by the CE 4 domain-containing deacetylase LcaB is required to confer a net positive charge to PIA and retain the polysaccharide on the bacterial cell wall (38). Similarly, PeA, a deacetylase in the *Pseudomonas aeruginosa* Pel exopolysaccharide biosynthetic operon, has also been reported to be required for the production of adhesive polysaccharide and biofilm formation in this organism (26). As with Agd3, PeA contains a small reductase fold and CE 4...
deacetylase domain separated by a region of predicted low complexity, as well as a β-sheet-rich region at the C terminus. While the role of the predicted reductase and C-terminal domains in PelA are unknown, the similarities between the domain arrangements within PelA and Agd3 suggest they likely mediate similar functions in the synthesis of Pel and GAG, respectively. PgaB, from *E. coli*, requires an adjacent domain, located C terminally to the CE 4 domain, for the deacylation of substrate (20). While there are domains other than CE 4 predicted in Agd3, their specific role in deacetylation is currently unknown. Finally, a protein containing the CE 4 domain, PssB, is required for the production of cell surface-bound extracellular polysaccharide in *Listeria monocytogenes* (39). Loss of PssB results in impaired bacterial cell aggregation and biofilm formation. Thus, postsynthesis modification of extracellular polysaccharides by deacetylation of hexosamine sugars seems to be a common theme used by diverse microorganisms in the formation of biofilms (40).

Although these polysaccharide systems display many similarities, several lines of evidence suggest that these biosynthetic pathways are the product of convergent, rather than divergent, evolution from a common ancestor of both fungi and bacteria. First, the polysaccharide composition and linkages of the exopolysaccharide product of these systems differ markedly between organisms. Although the composition of Pel is unknown, PIA/PNAG is composed of 1,6-linked GlcNAc residues; the exopolysaccharide of *L. monocytogenes* is composed of a 1,4-linked N-acetylmannosamine chain decorated with a terminal α-1,6-linked GlcNAc residue.

Deacetylation of *A. fumigatus* Exopolysaccharide

FIG 9 Agd3 orthologs are found within a wide range of *Ascomycetes* species. A gene tree is shown categorized by taxonomic class of Agd3 orthologs found in the 28 fungal species that possess the GAG gene cluster. The gene tree was built by aligning and trimming Agd3 ortholog sequences, followed by maximum likelihood phylogenetic analysis with 100 bootstrap replicates. The outgroup was rooted with sequences of *Arthrobasidium* species. The scale bar is the genetic distance representing amino acid substitutions per site.

FIG 10 *Trichosporon asahii* produces a GAG-like exopolysaccharide. Hyphae of *T. asahii* were stained with FITC-tagged soybean agglutinin (SBA) lectin binding for the detection of GalNAc-rich extracellular polysaccharide.
linked galactose; and GAG is a heteropolymer of α-1,4-linked galactose and GalNAc. Further, there is little sequence similarity between these bacterial and fungal enzymes beyond the conserved enzymatic domains themselves.

While it is possible that the GAG gene cluster arose de novo in fungi, it is also possible that these genes were acquired by horizontal gene transfer by a common ancestor of the Pezizomycotina that may have originated in bacteria or another organism. The clustered consecutive organization of the GAG biosynthetic genes is reminiscent of a bacterial operon and is relatively unusual in fungi. Fungal gene clusters have been best described as encoding enzymes required for the biosynthesis of secondary metabolites. Horizontal gene transfer of these secondary metabolite clusters has been suggested as a mechanism for their acquisition (41). The observations that T. asahii is the only basidiomycete found to contain the GAG gene cluster and that the T. asahii Agd3 orthologs with strong bootstrap support within the Sordariomycetes sequences in the gene tree are very strong evidence for a horizontal gene transfer event from an ascomycete species belonging to the Sordariomycetes class into T. asahii or its ancestor. Furthermore, the homology of T. asahii Agd3 and overall synteny of the cluster genes with other ascomycetes strongly suggest that the entire cluster was acquired in T. asahii or its parental lineages as a single horizontal transfer event. Although sequence analysis did not reveal any clear evidence of a specific mechanism and integration of the gene transfer, it is possible that it was mediated by a MULE domain-containing transposase, which is found near the boundary of the cluster in T. asahii. As genomes of more fungal species are made available and queried, a better picture of the timing of this exchange and overall evolution of this gene cluster will likely become clearer.

Interestingly, many of the species containing the GAG gene cluster are known plant pathogens, while no primary animal-associated fungi, such as the dermatophytes or Onygenales fungi, contain a homolog of the gene. Whether the ability to produce GAG confers an advantage to colonize and invade plant hosts and whether this selection pressure resulted in the convergent evolution of cationic polysaccharides in microorganisms remains to be determined. The observation that bacterial and fungal pathogens have developed similar strategies to produce cationic exopolysaccharide might suggest that these polymers facilitate pathogenicity. However, as humans are dead-end hosts for Aspergillus species, it is likely that the pressures responsible for the development of GAG were found in the soil or other natural environment. A shared requirement for adherence to substrates and resistance to environmental stresses through the production of an adherent layer of exopolysaccharide may underlie the development of exopolysaccharide synthesis by all of these microorganisms. Future studies examining the role of exopolysaccharide in fungal and bacterial growth in nonhuman environments may shed light on the factors leading to the evolution of these glycans in these varied microorganisms.

Deacetylation of GalNAc by Agd3 is required for A. fumigatus to produce cell wall-bound GAG and for mediating the phenotypes that have been associated with cell-wall-associated GAG. Further, the mixed mutant biofilm studies reported here would suggest that, as with Gram-positive bacterial exopolysaccharide, deacetylation occurs in the extracellular space. This observation suggests that inhibition of GAG deacetylation may represent an attractive antifungal target since intracellular penetration of a candidate deacetylase inhibitor would not be required for activity. Deacetylation of GalNAc within human cells has not been reported, suggesting that antifungal specificity may be achievable. As loss of GAG deacetylation has no direct effect on fungal viability, such a strategy would be best paired with a conventional antifungal agent.

Although a previous report did not identify the presence of deacetylated GalNAc within secreted GAG from A. fumigatus (3), compositional analysis of GAG from Aspergillus niger and Aspergillus parasiticus has reported the presence of GalN in preparations from these species (42, 43). Because deacetylated GAG adheres avidly to glass and plastics, it is possible that during the purification of A. fumigatus GAG, the preparation was enriched for fully acetylated, nonadherent polysaccharide. Alternately, it is possible that deacetylation varies under different growth conditions as reported in other species (44–46). Further experiments are required to test these possibilities.

In conclusion, we report that deacetylation of GAG by Agd3 is essential for the function of this exopolysaccharide, including adherence of GAG to the fungal cell wall and other substrates. These experiments shed light on the molecular mechanisms underlying GAG-mediated adhesion and draw important parallels between this glycan and bacterial exopolysaccharides. Finally, these data suggest the exciting possibility that targeting deacetylation of GAG may represent a promising antifungal strategy.

MATERIALS AND METHODS

Fungal strains and growth conditions. Aspergillus fumigatus strain AF293 (a generous gift from P. Magee, University of Minnesota, St. Paul, MN) was used as the parent strain for all genetic manipulations and experimental controls. Unless otherwise noted, all A. fumigatus strains were maintained on yeast extract-peptone-dextrose (YPD) agar (Fisher Scientific) at 37°C. For growth in liquid culture, Brian medium (3) or phenol-free RPMI 1640 (Wisent) was used as indicated in the figures at 37°C and 5% CO₂ incubation, unless otherwise noted. A clinical isolate of Trichosporon asahii was obtained from the McGill University Health Centre, Montreal, Quebec, Canada, and maintained on potato-dextrose medium at 30°C.

Molecular and genetic manipulations. A split marker, double homologous recombination approach was used to generate the Δagd3 mutant (47). Sequences flanking the open reading frame (ORF) of the agd3 gene were amplified from the genomic DNA using PCR (see Table S2 in the supplemental material). The resulting PCR fragments were then cloned into the entry vector pENTR-D-TOPO (Gateway, Inc.) and then recombined with previously described destination plasmids, pHY and pYG, containing the hygromycin split marker resistance cassettes (2). Target DNA was amplified by PCR from the recombined plasmids and then used for transformation of A. fumigatus as previously described (47).

To generate the Δagd3::agd3 complemented strain, the agd3 open reading frame and flanking sequences (680 bp downstream and 560 bp upstream) were amplified by PCR using the Sgf-compl-fow (compl stands for complemented, and fow stands for forward) and Asc-compl-rev (rev stands for reverse) primers, respectively (see Table S2 in the supplemental material). The resulting PCR fragment was cloned into pTAPA, a newly designed destination plasmid containing the phleomycin selection marker. The resulting pTAPA::agd3 plasmid contained the following sequence: Pagd3::ORFagd3::Tyc1::ORFPbio::Phia1::Trpc1. Two fragments for transformation were amplified by PCR from the pTAPA::agd3 plasmid using Sgf-compl-fow and LE4 primers and BL4 and Asc-compl-rev primers. Phleomycin-resistant transformants were selected, and correct integration at the native locus was verified by genomic PCR. Restoration of agd3 mRNA expression was confirmed by real-time reverse transcription-PCR (RT-PCR) (Table S2).

To generate the agd3::sfp mutant, the ORF of agd3 was amplified using

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primers Agel-agd-end-fow and EcoRV-agd-end-rev (see Table S2 in the supplemental material), cloned upstream of the rfp ORF in the pRFPy plasmid (17), and digested by Agel and EcoRV, which excised the gpdA promoter region on the plasmid. Amplification of the linearized plasmid produced the (agd3-end-ORF)::(RFP)::(TrpC)::(3′-half of HYG cassette) PCR fragment. From the agd3 terminator on the pRFPy plasmid, a 735-bp PCR product was made, using BsrGI-agd-Ter-fow and SacI-agd-Ter-rev. This product was cloned upstream of the hygromycin (HYG) cassette in the pAN7.1 plasmid (2), digested by BsrGI and SacI, resulting in an amplified PCR fragment (5′-half of HYG cassette)::(agd3-Tr). A. fumigatus AF293 was then transformed with this construct (47). All integrations were of agd3, and integration of the hygromycin resistance was verified by genomic PCR confirmed using real-time RT-PCR (see Table S2 for relevant primers) (2).

**Gene expression studies.** Expression of the galactosaminogalactan (GAG) cluster genes AFUA_3G07910 (age3), AFUA_3G07900 (sph3), AFUA_3G07890 (ega3), and AFUA_3G07860 (ega3) in the Δagd3 mutant was compared to the wild-type AF293. The fungal strains indicated in the figures were grown for 18 h at 37°C under various growth conditions including phenol-free RPMI 1640 (Wisent) buffered with morpholinepropanesulfonic acid (MOPS) (Bioshop, Inc.) at pH 5.4 and pH 7.0, Brian medium (3), or Aspergillus medium (2) with iron supplementation (NMR) detection of the primary amine group, dried GAG samples and purified as previously described (17). For chemical analysis to quantify polysaccharides, GAG samples were reconstituted in deuterated water, and cold deuterium chloride was used to negatively charge CM Sephadex beads (Sigma Aldrich, Inc.). The fungal strains indicated in the figures were grown in RPMI 1640 in 50-ml centrifuge tubes at 37°C and 5% CO2, for 18 to 24 h. Reconstituted sterile beads were added to the tubes containing pregrown fungi, and the tubes were then shaken for 10 min at 200 rpm. Supernatant samples were taken, and the number of unbound beads was counted by using a light microscope.

**Hyphal surf ace c h aracterization.** For biofilm adherence assays, fungi were grown for 24 h in BHI medium on tissue culture-treated six-well polystyrene plates. Nonadherent fungi were removed by vigorous agitation and washing. Adherent biofilms were visualized by crystal violet staining (53). Cell wall topography was examined using scanning electron microscopy as previously described (2). For soybean agglutinin (SBA) or recombinant Fc-dectin-1 binding to the surface of hyphae, strains were grown to 7 to 9 h on poly-D-lysine-coated coverslips (BD Biosciences, Inc.), then fixed, and stained with either fluorescein-tagged SBA or recombinant Fc-dectin-1 (2). For all microscopy experimentation, strains were grown in phenol red-free RPMI 1640. The susceptibility of strains to cell wall-active antifungals was determined as previously described (2). Bright-field images of mycelia were captured at a magnification of ×200 (Infinity 2 camera; Lumenera Inc.).

**Agd3 localization studies.** To determine the localization of Agd3 by fluorescence microscopy, the Δagd3-rfp mutant was grown on glass coverslips in RPMI 1640 for 9 to 12 h at 37°C and 5% CO2, fixed with 4% paraformaldehyde, stained with anti-red fluorescent protein (anti-RFP) IgG antibody (Abcam, Inc.), and detected with fluorescein-tagged anti-IgG antibody (Abcam, Inc.). The cells were then mounted on microscope slides and imaged by confocal microscopy as previously described (2). For RFP-tagged Agd3 (Agd3-RFP) immunoblotting, mycelia of the wild-type and Agd3-RFP strains were grown at 37°C for 24 h under shaking conditions, and culture supernatants were recovered by filtration. The biomass was crushed under liquid nitrogen, homogenized, and incubated for 1 h at 4°C in the presence of protease inhibitors (Bioshop, Inc.) and Triton X-100 at a final concentration of 2% (Bioshop, Inc.) prior to lyophilization. Protease inhibitors (Bioshop, Inc.) were added to the culture supernatant, lyophilized, and stored at −20°C. Detection of Agd3-RFP was performed by Western blotting using rat anti-RFP antibody (Chromotek, Inc.) and horseradish peroxidase (HRP)-conjugated donkey anti-rat IgG (Chromotek, Inc.).

**Polysaccharide analysis.** GAG was extracted by ethanol precipitation and purified as previously described (17). For chemical analysis to quantify the presence of primary amines in GAG, colorimetric detection using trinitrobenzene sulfonate (TNBS) (Fishier Scientific, Inc.) was performed following the manufacturer’s instructions. For nuclear magnetic resonance (NMR) detection of the primary amine group, dried GAG samples were reconstituted in deuterated water, and cold deuterium chloride was added to dissolve the samples. 1H heteronuclear multiple quantum correlation (HMOC) NMR experiments were conducted on a Varian INOVA 500-MHz spectrometer with a 3-mm gradient probe. Assignment of spectra was performed using Bruker Topspin version 3.1 program for spectrum visualization.
performed using Pinnacle v1.0.3.0 (Optyx Technologies) and the integrated counts from MS1 extracted ion currents specific to peptides in RFP and Agd3 verified by MS-MS.

**GAG enzyme immunoassay (EIA).** Culture filtrates from wild-type, Δagd3 mutant, or Δagd3 mutant strains alone or in combination were incubated for 1 h in a high-binding enzyme-linked immunosorbent assay (ELISA) plate (Nunclon Inc.). The wells were then washed with washing buffer, incubated for 1 h on a rotator with anti-GAG antibody (a kind gift from Jean-Paul Latgé, Institut Pasteur, Paris, France), then washed, incubated with anti-IgG HRP-tagged secondary antibody (Jackson Laboratories, Inc.), and developed by adding HRP substrate solution (Clontech, Inc.).

**Virulence studies.** Male BALB/c mice, 5 to 6 weeks old, were infected with the fungal strain indicated in the figures or sham infected with phosphate-buffered saline (PBS) using an aerosol chamber as previously described (30). Mice were immunosuppressed with 250 mg of cortisone acetate (Sigma-Aldrich) per kg of body weight by subcutaneous injection on days 2 and 3 and with 250 mg of cyclophosphamide (Western Medical Supply, Inc.) per kg intraaperitoneally on day 2 and 200 mg/kg on day 3 relative to conidial challenge (30). To prevent bacterial infection, enrofloxacin was added to the drinking water (Baytril, Inc.). Mice were monitored for a period of 2 weeks for signs of illness, and moribund animals were euthanized.

For pulmonary fungal burden determination, eight 5- to 6-week-old immunosuppressed male BALB/c mice were infected endotracheally with the fungal strains indicated in the figures as previously described (54). At day 4 postinfection, the lungs were harvested and either homogenized for fungal burden determination or fixed in formalin for histopathology. Fungal burden in lung homogenates was quantified by PCR, or the relative content of galactomannan was determined as previously described (2). To assess the extent of lung injury, bronchoalveolar lavage fluid samples from infected animals were collected prior to harvesting the lungs, and the activity of lactose dehydrogenase was determined per the manufacturer’s instructions (CytoTox96 nonradioactive cytotoxicity assay kit; Promega). For pulmonary histopathology examination, sections of lung were stained with periodic acid–Schiff (PAS) and digitally scanned at the Institute for Research in Immunology and Cancer (Université de Montréal). Scanned images were viewed, analyzed, and captured using ObjectView (Objective Pathology Services). All procedures involving mice were approved by the Los Angeles Biomedical Research Institute Animal Use and Care Committee and the McGill University Animal Use and Care Committee.

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at http://mbio.asm.orglookup/suppl/doi:10.1128/mBio.00252-16/-/DCSupplemental.

Figure S1, TIF file, 2 MB. Figure S2, TIF file, 2 MB. Figure S3, TIF file, 1.5 MB. Figure S4, TIF file, 2 MB. Figure S5, TIF file, 2.4 MB. Table S1, XLSX file, 0.04 MB. Table S2, XLSX file, 0.04 MB.

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**REFERENCES**


